

## SPECIAL ARTICLES

# *The functional matrix hypothesis revisited. 4. The epigenetic antithesis and the resolving synthesis*

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In two interrelated articles, the current revision of the functional matrix hypothesis extends to a reconsideration of the relative roles of genomic and of epigenetic processes and mechanisms in the regulation (control, causation) of craniofacial growth and development. The dialectical method was chosen to analyze this matter, because it explicitly provides for the fuller presentation of a genomic thesis, an epigenetic antithesis, and a resolving synthesis. The later two are presented here, where the synthesis suggests that both genomic and epigenetic factors are necessary causes, that neither alone is also a sufficient cause, and that only the two, interacting together, furnish both the necessary and sufficient cause(s) of ontogenesis. This article also provides a comprehensive bibliography that introduces the several new, and still evolving, disciplines that may provide alternative viewpoints capable of resolving this continuing controversy; repetition of the present theoretical bases for the arguments on both sides of these questions seems nonproductive. In their place, it is suggested that the group of disciplines, broadly termed Complexity, would most likely amply repay deeper consideration and application in the study of ontogenesis. (*Am J Orthod Dentofac Orthop* 1997;112:410-7.)

**I**t is a fallacy that the genome, the totality of DNA molecules, is the main repository for developmental information; i.e. that there exists a genetic program, or blueprint, theoretically capable of creating an entire organism.”<sup>98</sup>

### **Biological Mechanisms and Processes Defined**

This article continues the dialectical analysis of the roles of genomic and epigenetic processes and mechanisms in the control of craniofacial growth and development. Previously a genomic thesis was outlined and several critical terms were defined.<sup>99</sup> The dialectic process concludes here with an epigenetic antithesis and a resolving synthesis, following two additional definitions: (1) A process is a series of actions or operations that lead toward a particular result. (2) A mechanism is the fundamental physical or chemical process(es) involved in, or responsible for, an action, reaction, or other natural phenomenon.<sup>100</sup> That is, mechanisms underlie processes. For example, loading a femur is an epigenetic process: the possible resultant modification(s)

of bone cell DNA (for example by methylation<sup>101,102</sup>), or of chondrocytic DNA (for example as reflected in differential regulation of biosynthetic pathways<sup>103</sup>), are epigenetic mechanisms. Similarly, the specific steps of the activation and deactivation of appropriate portions of the bone cell genome, associated with the trio of possible osteoblastic responses to loading (deposition, resorption, or maintenance of bone tissue) are further examples of epigenetic mechanisms that control the genome. In this sense, the original versions of the functional matrix hypothesis (FMH) described only epigenetic processes,<sup>4-8</sup> whereas recent revisions also described epigenetic mechanisms.<sup>9,10</sup> The fundamental correctness of earlier FMH descriptions is supported by more recent research.<sup>104,105</sup>

### **The Epigenetic Antithesis**

Some of the principal strengths of this antithesis come from precise definitions of what a gene is and is not. For example: (a) “gene. The unit of heredity: one or more nucleic acid sequences incorporating information necessary for the generation of a particular peptide or RNA product”<sup>106</sup>; and, (b) “enough is known about the genetic machinery . . . [to know] . . . that this is virtually the only kind of information which polynucleotide molecules are inherently capable of containing: nothing there

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at all about which proteins will be expressed in which cells at what time and in what quantities.”<sup>98</sup>

The genomic thesis is denied because it is both reductionist and molecular; that is, descriptions of the causation (control, regulation) of all hierarchically higher and structurally more complex morphogenetic processes are reduced to explanations of mechanisms at the molecular (DNA) level. For example, the genomic thesis of craniofacial ontogenesis passes directly from molecules to morphogenesis: directly from DNA molecules to adult gross morphology, ignoring the role(s) of the many epigenetic processes and mechanisms competent to control (regulate, cause) the large number of intervening, and increasingly more structurally complex, developmental stages<sup>13,18</sup> particularly, and there are additional similarly reductionist views of odontogenesis.<sup>17,22,60,107,108</sup>

The epigenetic antithesis, detailing both processes and mechanisms, is integrative,<sup>109</sup> seeking to clarify the causal chain between genome and phenotype. Its goal is to identify and describe comprehensively the series of initiating biological processes and their related underlying (biochemical, biophysical) responsive mechanisms that are effective at each hierarchical level of increasing structural and operational complexity.<sup>110</sup>

This article reviews some of the clinically significant epigenetic processes and mechanisms, existing at several organizational (structural, functional) levels, that regulate (direct, control, cause) cephalic and craniofacial (musculo-) skeletal morphogenesis.

### Craniofacial Epigenetics

“Broadly speaking, epigenetics refers to the entire series of interactions among cells and cell products which leads to morphogenesis and differentiation. Thus all cranial development is epigenetic, by definition.” This view is supported here,<sup>15,19,20,111</sup> despite continued expressions of genomic regulation of craniofacial morphogenesis.<sup>13,14</sup>

As previously noted,<sup>99</sup> epigenetic factors include (1) all of the extrinsic, extraorganismal, macroenvironmental factors impinging on vital structures (for example, food, light, temperature), including mechanical loadings and electromagnetic fields, and (2) all of the intrinsic, intraorganismal, biophysical, biomechanical, biochemical, and bioelectric microenvironmental events occurring on, in, and between individual cells, extracellular materials, and cells and extracellular substances.

In terms of clinical orthodontics, and of the FMH, all therapy is applied epigenetics, and all

appliances (and most other therapies) act as prosthetic functional matrices. Clinical therapeutics includes a number of epigenetic processes, whose prior operations evoke a number of corresponding epigenetic mechanisms. These latter, in turn, underlie the observed processes of tissue adaptations by both skeletal units and functional matrices.

### Epigenetic Processes and Mechanisms

In craniofacial morphogenesis, more is known presently about processes than about mechanisms. Despite this, it is no longer sufficient to note, for example, that otherwise undescribed epigenetic processes of “intrauterine environment” can regulate fetal mandibular growth.<sup>112</sup> The future aim must be to elucidate the molecular, genomic, mechanisms<sup>101</sup> whose activation underlies the adaptive growth processes of the mandibular functional cranial components (that is, of the mandibular skeletal units and their related functional matrices).

### Loading

Many different epigenetic processes can evoke mechanisms capable of modifying DNA.<sup>113-116</sup> At clinically significant structural levels, physical loading is unquestionably of the greatest importance. “Among the numerous epigenetic factors influencing the vertebrate face is mechanical loading.”<sup>118</sup> It is useful to consider the epigenetic process of loading and some of the epigenetic mechanisms this process evokes.

*Loading per se.* Loads may be imposed at many structural levels. While clinical observations usually are macroscopic, the loadings act microscopically, at molecular and/or cellular levels.<sup>117</sup> Loadings are able to regulate several alternative molecular (cellular) synthetic pathways (mechanisms) of many tissues, including bone<sup>118</sup>; for example, the mechanical environment is important in maintaining the differentiated phenotype of bone cells.<sup>102</sup> It should be noted that loading may be dynamic (for example, muscle contraction) or static (that is, gravity); and to be effective, loads may increase, decrease, or remain constant.

Mechanical loading is known to influence gene expression.<sup>119,120</sup> Of clinical (and FMH) interest, extrinsic musculoskeletal loading can rapidly change (1) both articular cartilage intercellular molecular syntheses<sup>121</sup> and mineralization<sup>122</sup>; and (2) osteoblastic (skeletal unit) gene expression.<sup>123,124</sup> Epigenetic loading processes include gravitational variations that evoke unique mechanisms of molecular synthesis.<sup>125</sup>

*Extracellular matrix deformation.* Musculoskeletal tissue loading inevitably deforms an extracellular matrix (ECM) that is not developmentally inert. Rather, in several ways, ECM regulates the formation, development, and maintenance of its included cells that synthesize the ECM.<sup>126-129</sup> Further, ECM can regulate multicellular tissue morphogenesis<sup>130</sup> and contribute to genomic regulation of its enclosed cells.<sup>131</sup>

*Cell-shape changes.* Tissue loading can also alter cell shape. This inevitably deforms intracellular constituents, including the cytoskeleton.<sup>132-134</sup> The epigenetic process of changing cell shape invokes the epigenetic mechanisms of mechanotransduction of biophysical forces into genomic and morphogenetically regulatory signals.<sup>135-138</sup>

Cell-shape change processes can also activate several other epigenetic mechanisms, for example, stretch-activated ion channels in cartilage and other mechanically initiated cell-signaling mechanisms.<sup>139-142</sup> There is recent orthodontic interest in the cell-shape change of nonskeletal cells.<sup>143</sup>

Cell-shape change may lead to nuclear shape deformation. This, in turn, is a mechanism that can directly cause (regulate) a consequent alteration of the mechanisms of genomic activity.<sup>140</sup>

*Epigenetic cell signalling processes.* Several loading processes can regulate genomic expression. One, previously described, begins with cellular mechanoreception and mechanotransduction of the loading stimulus into an intercellular signal that undergoes parallel processing within a connected cellular network of bone cells.<sup>9,10</sup> The details of cell-signalling are reviewed extensively elsewhere.<sup>144</sup>

*Chains of intracellular molecular levers.* A second epigenetic cellular process begins with deformation of the ECM. This matrix has an epigenetic regulatory role in morphogenesis, by virtue of integrin molecules that physically interconnect the several molecular components of the intracellular (cytoskeletal) and the extracellular environment (for cartilage).<sup>145,127,128,146-148</sup> While the form (size and shape) of the cytoskeleton may be physically controlled by a broad spectrum of loadings,<sup>133,149</sup> it responds identically to all.<sup>150</sup>

The epigenetic mechanism evoked consists of a physical array of intracellular macromolecular chains, acting as levers, extending from the cell membrane to multiple specific sites on each chromosome.<sup>146</sup> The molecular chain acts as an information transfer system between the extracellular environment and the genome, transmitting signals generated by deformations of the ECM directly to

the intranuclear genome.<sup>9,10</sup> Indeed, such informational transfer between cells and ECM is dynamic, reciprocal, and continuous.<sup>151</sup>

*Other processes and mechanisms.* (1) DNA methylation is a potent epigenetic event. It is involved in many intracellular, extracellular, and intercellular mechanisms.<sup>101</sup> It can "introduce novel features of cellular function far removed from the classical Mendelian view of the gene, chromosome, and inheritance . . . with information flowing back to the DNA level and changing gene expression,"<sup>152,153</sup> the genome now being considered as a sophisticated response system and a carrier of information,<sup>154</sup> a system activated by several epigenetic processes and mechanisms.<sup>155</sup> (2) There are numerous examples of yet other processes and mechanisms of epigenetic regulation of the genome.<sup>113,115,156-159</sup> (3) In addition, it has been shown that (botanical) epigenetic factors can impose metastable inheritable changes in the plant genome,<sup>160-163</sup> a nontrivial matter not considered further here.

#### Epigenetic Regulation of Higher Structural Levels

In addition to the molecular and cellular processes and mechanisms noted, over a century ago the discipline of developmental mechanics (entwicklungsmechanik)<sup>85,86</sup> established that the epigenetic process of extrinsic loadings play a major role in the regulation of bone tissue and bone organ growth, development, and morphology.<sup>118,164-167</sup>

At the tissue level, there are several causal, strain-specific differences in bone tissue microstructure.<sup>168-171</sup> Closely similar epigenetic mechanisms and processes are observed in the adaptational responses of all connective tissues, including cartilage, to loading.<sup>164,165,172-175</sup>

At the organ level, the ability of the processes of motion and of articular function to regulate joint morphology is well-known<sup>176-178</sup>; and, of course, physical activity processes regulate organismal skeletal adaptational responses.<sup>179</sup> Other epigenetic processes affecting bone tissue include local vascular factors.<sup>180</sup>

*Regulation of functional matrices.* Periosteal functional matrices are under closely similar epigenetic control. Mechanical loads regulate skeletal muscle (periosteal functional matrix) phenotype<sup>181</sup>; and chronic muscle stimulation can change its phenotype.<sup>182-184</sup> Numerous studies establish the neurotrophic role of neural innervation in muscle genome regulation.<sup>185-188</sup> It remains only to note the truism that, for muscle as for bone, mechanical epigenetic factors, broadly termed function (or ex-

ercise) significantly control musculoskeletal growth, development,<sup>187,189,190</sup> and maintenance of structural and physiological attributes.<sup>191-193</sup>

### A Resolving Synthesis

“It seemed that the next minute they would discover a solution. Yet it was clear to both of them that the end was still far, far off, and that the hardest and most complicated part was only just beginning.”—Anton Chekov. *The Lady with the Dog*.

As the epigraph indicates, it is certain that no matter what arguments, theoretical constructs, and supporting experimental data are presented here, the prevailing tension between the genomic thesis and epigenetic antithesis will continue unabated. Nevertheless, a resolving synthesis will at least clarify the bases for continued discourse.

The fundamental argument of this resolving synthesis, based on an analysis of causation, was presented earlier,<sup>11</sup> and later amplified.<sup>99</sup> It argues that morphogenesis is regulated (controlled, caused) by the activity of both genomic and epigenetic processes and mechanisms. Both are necessary causes; neither alone are sufficient causes; and only their integrated activities provides the necessary and sufficient causes of growth and development. Genomic factors are considered as intrinsic and prior causes; epigenetic factors are considered as extrinsic and proximate causes. The data supporting this synthesis are provided here and above.<sup>99</sup>

It is acknowledged that the validity of this dialectic synthesis is significantly dependent on the validity of its epigenetic antithesis. In turn, a defensible epigenetic antithesis should convincingly suggest some process(es) and/or mechanism(s) that can regulate (direct, control, cause) morphogenesis. It is argued here that these are provided by the newly emerging disciplines of complexity.

### Complexity and self-organization

The theories of ontogeny and phylogeny currently are being significantly reinvigorated by the new and evolving science(s) of complexity that integrate topics from mathematics (for example, cellular automata, fractals, strange attractors), biology (for example, genetic algorithms, artificial life simulations, neural networks, emergence, adaptive systems, connectivity), and physics, while minimizing distinctions between them. Complexity theory (CT) also integrates specifically related topics in bioengineering and the computer sciences; for example, chaos, information, and hierarchical theories, fuzzy logic, as well as cyto(tissue)mechanics and molecular (nano)mechanics.<sup>194-212</sup>

Because epigenetic processes and mechanisms are best explained as examples of CT, a clearcut demonstration of the role of CT in craniofacial ontogeny, at some point, is both necessary and possible. But in this place only this brief, intuitive preview is possible. Because fairness to both the novelty and conceptual richness of CT requires a comprehensive presentation to make it generally intelligible, it will be substantively reviewed subsequently.

CT provides descriptions of the behavior of complex biological systems that exist as “ensembles” of several tissues and organs, and not as clusters of individual cells and extracellular substances. Such an ensemble (identical to a functional cranial component in the FMH) is termed here as a complex adaptive system (CAS), structurally arrayed as a vital continuum. This term is defined here as it is in the several analytical finite element methods (FEM) recently introduced into orthodontics and physical anthropology.<sup>213-221</sup>

CT provides compact, statistical descriptions of the collective growth behavior of such CAS continuity. During ontogeny, vital CAS exhibits the creation of robust, spontaneous, and emergent order.

An algorithm for control of such a CAS requires that it is able to alter itself in response to the (epigenetic) information produced by the system it is trying to control. In a CAS, minor changes in the epigenetic input can cause huge fluctuations in the morphological output.

CT, as it utilized information theory, assumed that a CAS processes information (both genomic and epigenetic) in a parallel, not a serial, manner.<sup>10</sup> Where most previous biological theories of development were based on the methods of deterministic (genomically predetermined), classical mechanics, information theory, and CT, are probabilistic (epigenetically self-organized and emergent), and are based on the methods of statistical mechanics. It is probable that ontogeny involves nonlinear processes and is not fully predictable; that is, growth and development, to a significant extent, exhibit both random behaviors and frequent perturbations. To clarify this point, note that previously most biological models were studied as if they were linear. That is, when their mathematical formulas were graphed they looked like straight lines. Linear systems are predictable: the calculus shows the changes in their state, and statistics (especially regression analysis) reduces their data to a line. However, CT makes it clear that most biological systems are nonlinear and are not most correctly described by these mathematical techniques; nonlinear formulations are necessary.

The highly ordered morphological properties of adult complex biological systems (for example, functional matrices and skeletal units) result from the operation of a series of spontaneous and self-organized ontogenetic processes and mechanisms.<sup>194,200</sup> Such emergent self-organizing events can create phenotypic variability under constant genetic and other extraorganismal epigenetic conditions.<sup>222</sup>

The operation of complexity can be suggested as follows. "Environmental factors thus play a decisive role in all ontogenetic processes. But it is the organism itself that, as an integrated system, dictates the nature of each and every developmental response . . . the living organism self-organizes on the basis of its own internal structuring, in continuous interaction with the environment in which it finds itself."<sup>113</sup>

## CONCLUSIONS

Integration of pertinent advances in biomedical and bioengineering permitted an ongoing revision of the functional matrix hypothesis. The first two articles in this series, by emphasizing the roles of a number of biophysical and biochemical factors in the regulation of morphogenesis, implicitly argued for the correctness of the fundamentally epigenetic thrust of the FMH. However, because the conceptual tension between hypotheses suggesting the regulatory primacy of either genomic (genetic) or of epigenetic factors and/or processes in morphogenesis continues unabated, it seemed useful to reevaluate this nontrivial matter, using the dialectical method of presenting a thesis, an antithesis, and a resolving synthesis as illustrated in these two interrelated articles.

I believe that the most appropriate conclusion permitted by the data bases at this time is to use the contemporary managerial phrase . . . "it is a win-win situation." Again, using a popular phrase, genomic and epigenetic processes are "apples and pears" More correctly, they are examples of totally differing types of causation—genomic formal cause and epigenetic efficient cause. Individually both are necessary causes, but neither are sufficient causes alone. Together they provide both the necessary and sufficient causes for the control (regulation) of morphogenesis. Nevertheless, epigenetic processes and events are the immediately proximate causes of development, and as such they are the primary agencies. The fuller demonstration of exactly how epigenetic events carry out their roles will be considered elsewhere in the context of a review of the implications of complexity theory for the functional matrix hypothesis.

## REFERENCES

- Jacob F. The logic of life. New York: Pantheon Books, 1973.
- Kessler DS, Melton DA. Vertebrate embryonic induction: mesodermal and neural patterning. *Science* 1994;266:596-604.
- Moss ML, Young R. A functional approach to craniology. *Am J Phys Anthropol* 1960;18:281-92.
- Moss ML. The functional matrix. In: Kraus B, Reidel R, editors. *Vistas in orthodontics*. Philadelphia: Lea & Febiger, 1962:85-98.
- Moss ML. The primacy of functional matrices in orofacial growth. *Trans Br Soc Study Orthodont and Dent Pract* 1968;19:65-73.
- Moss ML. Twenty years of functional cranial analysis. *Am J Orthodont* 1972;61:479-85.
- Moss ML, Salentijn L. The primary role of functional matrices in facial growth. *Am J Orthodont* 1969a;55:566-77.
- Moss ML, Salentijn L. The capsular matrix. *Am J Orthodont* 1969b;56:474-90.
- Moss ML. The functional matrix hypothesis revisited. 1. The role of mechanotransduction. *Am J Orthodont Dentofac Orthop* 1997;111:8-11.
- Moss ML. The functional matrix hypothesis revisited. 2. The role of an osseous connected cellular network. *Am J Orthodont Dentofac Orthop* 1997;112:221-6.
- Moss ML. Genetics, epigenetics, and causation. *Am J Orthodont* 1981;80:366-75.
- Johnston MC, Bronsky PT. Prenatal craniofacial development: new insights on normal and abnormal mechanisms. *Crit Rev Oral Biol Med* 1995;6:368-422.
- Dixon MJ. Positional cloning of a gene involved in the pathogenesis of Treacher Collins syndrome. *Nature Genetics* 1996;12:130-6.
- Winter RM. Whats in a face? *Nature Genetics* 1996;12:24-9.
- Herring SW. Epigenetic and functional influences on skull growth. In: Hanken J, Hall BK, editors. *The skull*. Chicago: University of Chicago Press 1993a;1:153-206.
- Dawkins R. *The selfish gene*. Oxford: Oxford University Press, 1976.
- Harold FM. Ionic and electrical dimensions of hyphal growth. In: Wessels JG, Meinhardt F, editors. *The Mycota*. Berlin: Springer Verlag 1994;1:89-107.
- Harold FM. From morphogenes to morphogenesis. *Microbiol* 1995;141:2765-78.
- Herring SW. Formation of the vertebrate face: epigenetic and functional influences. *Am Zool* 1993b;33:472-83.
- Hall BK. Epigenetic control in development and evolution. In: Goodwin BC, Holder N, Wylie CG, editors. *Development and evolution*. Cambridge: Cambridge University Press, 1983:353-79.
- Holliday R. DNA methylation and epigenetic mechanisms. *Cell Biophys* 1989;15:15-20.
- Holliday R. Epigenetics: an overview. *Dev Genet* 1994;16:453-7.
- Lewontin RC. The dream of the human genome. *New York Review of Books* 1992;39:31-40.
- Lewontin RC. *Biology as ideology. The doctrine of DNA*. New York: Harper-Collins, 1992.
- Lewontin RC, Rose S, Kamin LJ. *Not in our genes*. New York: Pantheon Books, 1984.
- Ho MW, Saunders PT. Beyond neo-Darwinism—an epigenetic approach to evolution. *J Theor Biol* 1979;78:573-91.
- Lovtrup S. *Epigenetics. A treatise on theoretical biology*. London: John Wiley & Sons, 1974.
- Brien P. *Le vivant: epigenese, evolution, epigenetique*. Brussels: Ed. de L'Universite de Bruxelles, 1974.
- Bleschschmidt E, Gasser RF. *Biokinetics and biodynamics of human differentiation*. Springfield: C. C. Thomas, 1978.
- Corner MA. Reciprocity of structure-function relations in developing neural networks: the Odyssey of a self-organizing brain through research fads, fallacies and prospects. *Prog Brain Res* 1994;102:3-31.
- Jablonska E, Lamb MJ. *Epigenetic inheritance and evolution*. Oxford: Oxford University Press, 1995.
- Holland JH. *Hidden order. How adaptation builds complexity*. Reading MA: Addison-Wesley, 1995.
- Bonner JT. *The evolution of complexity by means of natural selection*. Princeton: Princeton University Press, 1988.
- Edelman GM. *Topoly: an introduction to molecular embryology*. New York: Basic Books, 1988.
- Gell-Mann M. *The quark and the jaguar: adventures in the simple and the complex*. New York: Freeman, 1994.
- Holland JH. *Adaptation in natural and artificial systems: an introductory analysis for adaptation in machine and nature*. Amsterdam: North-Holland, 1992.
- Waldrop MM. *Complexity: the emerging science at the edge of order and chaos*. New York: Simon & Schuster, 1992.
- Bassingthwaite JB, Liebovitch JS, West BJ. *Fractal physiology*. New York: Oxford University Press, 1994.
- Grande L, Rieppel O. *Interpreting the hierarchy of nature*. San Diego: Academic Press, 1994.
- Kauffman SA. *The origins of order*. New York: Oxford University Press, 1993.
- Mitchell M. *An introduction to genetic algorithms*. Cambridge: MIT Press, 1996.
- Langton CG, editor. *Artificial life. An overview*. Cambridge: MIT Press, 1995.
- Salmivirta K, Gullberg D, Hirsch E, Altruda F, Ekblom P. Integrin subunit expression associated with epithelial-mesenchymal interactions during murine tooth development. *Dev Dyn* 1996;205:104-13.
- Huyseune A. Phenotypic plasticity in the lower pharyngeal jaw dentition of *Astatoreochromis alluadi* (Teleostei:Chichlidae). *Arch Oral Biol* 1995;40:1005-14.
- Meyer A. Ecological and evolutionary consequences of the trophic polymorphism in *Cichasoma citrillum* (Pices: Chichlidae). *Biol J Linn Soc* 1990;39:279-99.
- Vandewalle P, Huyseune A, Aerts P, Verraes W. The pharyngeal apparatus in teleost feeding. In: Bels V, Chardon M, Vandewalle P, editors. *Biomechanics of feeding invertebrates*. Berlin: Springer Verlag, 1994:59-92.

47. Slavkin H. Editorial. Genetic and epigenetic challenges in tooth development. *J Craniofac Genet Dev Biol* 1988;8:195-8.
48. Lodish H, Baltimore D, Berk A, Zipursky SL, Matsudaira P, Darnell J. *Molecular cell biology* 3rd ed. New York: Sci Am Books, WH Freeman and Company, 1995.
49. Alberts B, Bray D, Lewis J, Raff M, Roberts, Watson JD. *Molecular biology of the cell*. 2nd ed. New York: Garland Publ, 1989.
50. Bishop JE, Waldholz M. *The genome. The story of the most astonishing scientific adventure of our time. The attempt to map all the genes in the human body*. New York: Simon & Schuster, 1992.
51. Wills C. Exons, introns, and talking genes: the science behind the human genome project. New York: Basic Books, 1992.
52. Cotran RS, Kumar V, Robbins SL. *Pathologic basis of disease*. Philadelphia: WB Saunders, 1994.
53. Darnell J, Lodish H, Baltimore D. *Molecular cell biology*. 2nd ed. New York: Sci Am Books, 1990.
54. Morgan TH. *The theory of the gene*. New Haven: Yale University Press, 1926.
55. Sinnot EW, Dunn LC. *The principles of genetics*. New York: McGraw-Hill Book Company, 1939.
56. Kevles DJ. *In the name of eugenics: genetics and the uses of human heredity*. New York: Knopf, 1985.
57. Gould S. *The mismeasure of man*. New York: WW Norton, 1981.
58. Mayr E, Provine WB. *The evolutionary synthesis*. Cambridge: Harvard University Press, 1981.
59. Davis J. *Mapping the code: the human genome project and the choices of modern science*. New York: John Wiley, 1992.
60. Olson MV. A time to sequence. *Science* 1995;270:394-6.
61. Crystal RG. Transfer of genes to humans: early lessons and obstacles to success. *Science* 1995;270:404-9.
62. Mann CC. Behavioral genetics in transition. *Science* 1994;264:1686-9.
63. Bouchard TJ Jr. Genes, environment and personality. *Science* 1994;264:1700-1.
64. Crabbe JC, Belknap JK, Buck KJ. Genetic animal models of alcohol and drug abuse. *Science* 1994;264:1715-23.
65. Myers MP, Wager-Smith K, Wesley CS, Young WY, Segal A. Positional cloning and sequence analysis of the *Drosophila* clock gene, timeless... *Science* 1995;270:805-8.
66. Blum K, Cull JG, Braverman ER, Comings DE. Reward deficiency syndrome. *Am Sci* 1996;84:132-45.
67. Herrstein RJ, Murray C. *The Bell curve*. New York: The Free Press, 1994.
68. Reddi AH. Cartilage morphogenesis: the role of bone and cartilage morphogenetic proteins, homeobox genes and extracellular matrix. *Matrix Biol* 1994;14:599-606.
69. Atchley WR. Genetic and developmental aspects of variability in the mammalian mandible. 1993. In: Hanken J, Hall BK, editors. *The skull*. Chicago: University of Chicago Press, 1993:1:207-47.
70. Ignelzi MA, Liu Y-H, Maxson RE Jr, Sneed ML. Genetically engineered mice: tools to understand craniofacial development. *Crit Rev Oral Biol Med* 1995;6:181-201.
71. Ishiguro K. Genetic study of mandibular prognathism by multi variate analysis. *Jpn J Stomat Soc* 1972;38:58-72.
72. Lovell DP, Johnson FM. Quantitative genetic variation in the skeleton of the mouse. 1. Variation between inbred strains. *Genet Res Camb* 1983;42:169-82.
73. Festing M. A multivariate analysis of subline divergence in the shape of the mandible in C57BL/Gr mice. *Genet Res Camb* 1973;21:121-32.
74. Bailey DW. Genes that affect the shape of the murine mandible. Congenic strain analysis. *J Hered* 1985;76:107-14.
75. Bailey DW. Genes that affect the shape of the murine mandible. Recombinant-inbred strain analysis. *J Hered* 1986a;77:17-25.
76. Bailey DW. Mandibular-morphogenesis gene linked to the H-2 complex in mice. *J Craniofac Genet Dev Biol* 1986b;2 (Suppl):S33-9.
77. Lacombe D. Clinical dysmorphology beyond developmental genetics: recent advances in some human developmental genes. *Int Rev Genet* 1995;38:137-44.
78. Balme DM. 1992 Aristotle. *De Partibus Animalium I and De Generatione Animalium I*. Oxford: Clarendon Aristotle Series. Clarendon Press, 1992.
79. Boyd E. *Origins of the study of human growth*. Portland, OR: University of Oregon, School of Dentistry, 1980.
80. Ross,WD, Smith JA, editors. *Works of Aristotle*. Oxford: Oxford University Press, 1908-12.
81. Singer C. *A short history of anatomy and physiology from the Greeks to Harvey*. 2nd ed. New York: Dover Publ, 1957.
82. Vogl C, Atchley WR, Cowley D, Crenshaw P, Murphy JD, Pomp D. The epigenetic influence of growth hormone on skeletal development. *Growth Dev Aging* 1993;57:163-82.
83. Scandalios JG, ed. *Genomic responses to environmental stress*. San Diego: Academic Press, 1990.
84. Bernard C. *An introduction to the study of experimental medicine*. London: Macmillan, 1855 (translation 1927).
85. Counce SJ. Archives for developmental mechanics. W. Roux, editor (1894-1924). *Roux's Arch Dev Biol* 1994;204:79-92.
86. Churchill FB. Chabry, Roux and the experimental method in nineteenth-century embryology. In: Giere RN, Westfall RS, editors. *Foundations of the scientific method: the nineteenth century*. Bloomington: Indiana University Press, 1973.
87. Kauffman S. *The origins of order. Self-organization and selection in evolution*. Oxford: Oxford University Press, 1993.
88. Kauffman S. *At home in the universe. The search for the laws of self-organization and complexity*. Oxford: Oxford University Press, 1995.
89. Waldrop MM. *Complexity. The emerging science at the edge of order and chaos*. New York: Simon & Schuster, 1992.
90. Lewin R. *Complexity. Life at the edge of chaos*. New York: Macmillan, 1992.
91. Holland JH. *Hidden Order. How adaptation builds complexity*. Reading MA: Addison-Wesley, 1995.
92. Teasta B, Kier LB. Complex systems in drug research. 1. The chemical levels. *Complexity* 1996;1(4):29-36.
93. Rose G, Siebler M. Cooperative effects of neuronal assemblies. *Exp Brain Res* 1995;106:106-10.
94. Hopfield JJ. Neural networks and physical systems with emergent computational abilities. *Proc Natl Acad Sci USA* 1982;79:2254-8.
95. Segel LA. Grappling with complexity. *Complexity* 1995;1(2):18-25.
96. Wallace WA. *Causality and scientific explanation*. Ann Arbor: University of Michigan Press, 1974.
97. Muragaki Y, Mundlos S, Upton J, Olsen BR. Altered growth and branching patterns in synpolydactyly caused by mutations in HOXD13. *Science* 1996;272:548-51.
98. Corner MA. Reciprocity of structure-function relations in developing neural networks: the Odyssey of a self-organizing brain through research fads, fallacies and prospects. *Prog Brain Res* 1994;102:3-31.
99. Moss ML. The functional matrix hypothesis revisited. 3. The genomic thesis. *Am J Orthod Dentofac Orthop* 1997;112:338-42.
100. Gove PB, editor. *Webster's seventh new collegiate dictionary*. Springfield MA: GC Merriam Co, 1970.
101. Hodgkin J. Epigenetics and the maintenance of gene activity states in *Caenorhabditis elegans*. *Dev Genet* 1994;15:471-77.
102. Roelofsens J, Klein-Nulend J, Burger EH. Mechanical stimulation by intermittent hydrostatic compression promotes bone-specific gene expression in vitro. *J Biomech* 1995;28:1493-503.
103. Kim Y-J, Grodzinsky AJ, Ploas AHK. Compression of cartilage results in differential effects on biosynthetic pathways for aggrecan link protein and hyaluronan. *Arch Biochem Biophys* 1996;328:331-40.
104. Civitelli R. Cell-cell communication in bone. *Calcif Tissue Int* 1995;56(Suppl 1):S29-S31.
105. Duncan RL, Turner CH. Mechanotransduction and the functional response of bone to mechanical strain. *Calcif Tissue Int* 1995;57:344-58.
106. Meier AE, editor. A is for... gene. *Sci Med* 1996;3:72.
107. Vernon RB, Gage EH. Between molecules and morphology. Extracellular matrix and the creation of vascular form. *Am J Pathol* 1995;147:873-83.
108. Sunohara M, Tanzawa H, Kaneko Y, Fuse A, Sate K. Expression patterns of Raf-1 suggest multiple roles in tooth development. *Calcif Tissue Int* 1996;58:60-4.
109. Noble D, Boyd CAR. The challenge of integrative physiology. In: Noble D, Boyd CAR, editors. *The logic of life. The challenge of integrative physiology*. Oxford: Oxford University Press, 1993:1-13.
110. Green DG. Emergent behavior in biological systems. In: Green DG, Bossmaier TJ, editors. *From biology to computation*. Amsterdam: IOS Press, 1993:424-35.
111. Ruff CB, Trinkhaus E, Walker A, Larsen CS. Postcranial robusticity in *Homo*. 1. Temporal trends, mechanical interpretation. *Am J Phys Anthropol* 1993;91:21-53.
112. Sasaki Y, Nonaka K, Nakata M. The effects of four strains of recipients on the intrauterine growth of the mandible in mouse fetuses. *J Craniofac Genet Dev Biol* 1994;14:118-23.
113. Latham KE, McGrath J, Solter D. Mechanistic and developmental aspects of genetic imprinting in mammals. *Int Rev Cytol* 1995;160:53-98.
114. Jorgensen R. Developmental significance of epigenetic impositions on the plant genome: a paragenetic function for chromosomes. *Dev Genet* 1994;15:523-32.
115. Moehrl A, Paro R. Spreading the silence: epigenetic transcriptional regulation during *Drosophila* development. *Dev Genet* 1994;15:478-84.
116. Meinhardt H. Pattern formation and the activation of particular genes. In: Goldbeter A, editor. *Cell-cell signaling: from experiment to theoretical models*. London: Academic Press, 1989.
117. Davies PF, Brand RA. editors. *Cell mechanics*. 1995. *J Biomech* 1995;28:1411-569.
118. Herring SW. Development of functional interactions between skeletal and muscular systems. In: Hall BK, editor. *Bone*. Boca Raton: Academic Press 1994;9:165-91.
119. Sandy JR, Fardale RW, Meikle MC. Recent advances in understanding mechanically induced bone remodeling and their relevance to orthodontic theory and practice. *Am J Orthod Dentofac Orthop* 1993;103:21-22.
120. Nebe B, Rychly J, Knopp A, Bohn W. Mechanical induction of (beta)1-integrin-mediated calcium signaling in a hepatocyte cell line. *Exp Cell Res* 1995;218:479-84.
121. Visser NA, Vankampen GPJ, Dekoning MHMT, Vanderkorst JK. The effects of

- loading on the synthesis of biglycan and decorin in intact mature articular cartilage in vitro. *Connect Tissue Res* 1994;30:241-50.
122. van't Veen SJ, Hagen JW, van Ginkel FC, Prahl-Andersen P, Burger EH. Intermittent compression stimulates cartilage mineralization. *Bone* 1995;17:461-5.
  123. Raab-Cullen DM, Thiede DN, Petersen DN, Kimmel DB, Recker RR. Mechanical loading stimulates rapid changes in periosteal gene expression. *Calcif Tissue Int* 1994;55:473-8.
  124. Stanford Cm, Stevens JW, Brand RA. Cellular deformation reversibly depresses rt-pcr detectable levels of bone-related mRNA. *J Biomech* 1995;28:1419-27.
  125. Guignandon A, Vico L, Alexandre C, Lafarge-Proust M-H. Shape changes of osteoblastic cells under gravitational variations during parabolic flight—relationship with PGE<sub>2</sub> synthesis. *Cell Struct Funct* 1995;20:369-75.
  126. Rodan GA, Rodan SB. The cells of bone. In: Riggs BL, Melton LJ, editors. *Osteoporosis: etiology, diagnosis and management*. 2nd ed. Philadelphia: Lippincott-Raven Publ;1995:1-39.
  127. Golombick T, Dajce DR, Bezwoda WR. Extracellular matrix interactions: 1. Production of extracellular matrix with attachment and growth-sustaining functions by UWOV2 ovarian cancer cells growing in protein-free conditions in vitro. *Cell Dev Biol* 1995;31:387-95.
  128. Yamada KM, Miyamoto S. Integrin transmembrane signaling and cytoskeletal control. *Curr Opin Cell Biol* 1995;7:681-9.
  129. Ekblom P. Extracellular matrix in animal development—an introduction. *Experientia* 1995;51:851-2.
  130. Gumbiner BM. Cell adhesion: the molecular basis of tissue architecture and morphogenesis. *Cell* 1996;84:345-57.
  131. Roskelley CD, Bissell MJ. Dynamic reciprocity revisited: a continuous, bidirectional flow of information between cells and the extracellular matrix regulates mammary epithelial cell function. *Biochem Cell Biol* 1995;73:391-7.
  132. Carvalho RS, Scott JE, Suga DM, Yen EHK. Stimulation of signal transduction pathways in osteoblasts by mechanical strain potentiated by parathyroid hormone. *J Bone Miner Res* 1994;9:999-1011.
  133. Wang N, Ingber DE. Control of cytoskeletal mechanics by extracellular matrix, cell shape and mechanical tension. *Biophys J* 1994;66:2181-9.
  134. Iwig M, Czeslick E, Muller A, Gruner M. Growth regulation by cell shape alteration and organization of the cytoskeleton. *Europ J Cell Biol* 1995;67:147-57.
  135. Gordon R. Mechanical engineering of the cytoskeleton in developmental biology. *Int Rev Cytol* 1994;150:1-150.
  136. Ingber DE, Dike L, Karp S, Liley H. Cellular tensegrity: exploring how the mechanical changes in the cytoskeleton regulate cell growth, migration and tissue pattern during morphogenesis. *Int Rev Cytol* 1994;150:173-220.
  137. Belousov LV, Savaliev SV, Naumidi II, Novoselov VV. Mechanical stresses in embryonic tissues: patterns, morphogenetic role and involvement in regulatory feedback. *Int Rev Cytol* 1994;150:1-33.
  138. Goodwin BC. What causes morphogenesis? *Bioessays* 1985;3:35-6.
  139. Wright M, Jobanputra PM, Bavington C, Salter DM, Nuki G. Effects of intermittent pressure-induced strain on the electrophysiology of cultured human chondrocytes: evidence for the presence of stretch-activated membrane ion channels. *Clin Sci* 1996;90:61-71.
  140. Matyas J, Edwards P, Miniaci A, Shrive N, Wilson J, Bray R, Frank C. Ligament tension affects nuclear shape in situ: an in vitro study. *Connect Tissue Res* 1994;31:45-53.
  141. Duncan RL, Hruska KA. Chronic, intermittent loading alters mechanosensitive channel characteristics in osteoblast-like cells. *Am J Physiol* 1994;267:F909-16.
  142. Guilak F, Ratcliffe A, Mow VC. Chondrocyte deformation and local tissue strain in articular cartilage: a confocal microscopy study. *J Orthop Res* 1995;13:410-21.
  143. Norton LA, Andersen KL, Arenholt-Bindslev D, Andersen L, Melsen B. A methodical study of shape change in human oral cells perturbed by a simulated orthodontic strain in vitro. *Arch Oral Biol* 1995;40:863-72.
  144. Banes AJ, Tsuzaki M, Yamamoto J, Fischer T, Brigman B, Brown T, Miller L. Mechanoreception at the cellular level: the detection, interpretation, and diversity of responses to mechanical signals. *Biochem Cell Biol* 1995;73:349-65.
  145. Holmwall K, Camper L, Johansson S, Kimura JH, Lundgren-Akerlund E. Chondrocyte and chondrosarcoma cell integrins with affinity for collagen type II and their response to mechanical stress. *Exp Cell Res* 1995;221:496-503.
  146. Puck TT, Krystosek A. Role of the cytoskeleton in genome regulation and cancer. *Int Rev Cytol* 1992;132:75-108.
  147. Wilson E, Sudhir K, Ives HE. Mechanical strain of rat vascular smooth muscle cells is sensed by specific extracellular matrix/integrin interactions. *J Clin Invest* 1995;96:2364-72.
  148. Couchman JR, Woods A. Transmembrane signaling generated by cell-extracellular matrix interactions. *Kidney Int* 1995;47(Suppl 49):S8-12.
  149. Ingber DE. The riddle of morphogenesis: a question of solution chemistry molecular cell engineering? *Cell* 1993;75:1249-52.
  150. Haskin C, Cameron I. Physiological levels of hydrostatic pressure alter morphology and organization of cytoskeletal and adhesion proteins in MG-63 osteosarcoma cells. *Biochem Cell Biol* 1993;71:27-35.
  151. Roskelley CD, Srebrow A, Bissell MJ. A hierarchy of ECM-mediated signalling regulates tissue-specific gene expression. *Curr Opin Cell Biol* 1995;7:736-47.
  152. Holliday R. Mechanisms for the control of gene activity during development. *Biol Rev* 1990;65:431-71.
  153. Holliday R. DNA methylation, epigenetic inheritance. *Philos Trans R Soc Lond B Biol Sci* 1990;326:329-38.
  154. Jablonka E, Lamb MJ. *Epigenetic inheritance and evolution*. Oxford: Oxford University Press, 1995.
  155. Janousek B, Siroky J, Vyskot B. Epigenetic control of sexual phenotype in a dioecious plant, *Melandrium album*. *Mol Gen Genet* 1996;250:483-90.
  156. Latham KE. Strain-specific differences in mouse oocytes and their contribution to epigenetic inheritance. *Development* 1994;120:3419-26.
  157. Bestor TH, Chandler VL, Feinberg AP. Epigenetic effects in eukaryotic gene expression. *Dev Genet* 1994;15:458-62.
  158. Wolpert L. Positional information and pattern formation in development. *Dev Genet* 1994;15:485-90.
  159. MacLeod MC. A possible role in chemical carcinogenesis for epigenetic, heritable changes in gene expression. *Mol Carcinog* 1996;15:241-50.
  160. Jorgensen R. Developmental significance of epigenetic impositions on the plant genome: a paragenetic function for chromosomes. *Dev Genet* 1994;15:523-32.
  161. Mikula BC. Environmental programming of heritable epigenetic changes in paramutated r-gene expression using temperature and light at a specific stage of early development in maize seedlings. *Genetics* 1995;140:1379-87.
  162. Clifton KH. Comments on the evidence in support of the epigenetic nature of radiogenic initiation. *Mutat Res* 1996;350:77-80.
  163. McClintock B. The control of gene action in maize. *Brookhaven Symp Biol* 1965;18:162-84.
  164. Biewener AA, Bertram JEA. Mechanical loading and bone growth in vivo. In: Hall BK, editor. *Bone. Bone growth-B*. Boca Raton: CRC Press, 1993;7:1-36.
  165. Burger EH, Veldhuijzen JP. Influence of mechanical factors on bone formation, resorption and growth in vitro. In: Hall BK, editor. *Bone. Bone growth-B*. Boca Raton: CRC Press, 1993;7:37-56.
  166. Schaffer JL, Rizen M, L'Italien G, Benbrahim A, Megerman J. Device for the application of a dynamic biaxially uniform and isotropic strain to a flexible cell culture membrane. *J Orthop Res* 1994;12:709-19.
  167. Skedros JG, Mason MW, Bloebaum RD. Differences in osteonal micromorphology between tensile and compressive cortices of a bending skeletal system: indications of potential strain-specific differences in bone microstructure. *Anat Rec* 1994;239:405-13.
  168. McMahon JM, Boyde A, Bromage TG. Pattern of collagen fiber orientation in ovine calcaneal shaft and its relation to locomotor-induced strain. *Anat Rec* 1995;242:147-58.
  169. Raspanti M, Guizzardi S, Stocchi R, Ruggeri A. Different fibrillar architectures coexisting in Haversian bone. *Ital J Anat Embryol* 1995;100(Suppl 1):103-12.
  170. Petryl M, Hert J, Fiala P. Spatial organization of the haversian bone in man. *J Biomech* 1995;29:161-9.
  171. Suh J-K, Li Z, Woo L-Y. Dynamic behavior of a biphasic cartilage model under cyclic compressive loading. *J Biomech* 1995;28:357-64.
  172. Kantomaa T, Tuominen M, Pirttiniemi P. Effect of mechanical forces on chondrocyte maturation and differentiation in the mandibular condyle of the rat. *J Dent Res* 1994;73:1150-6.
  173. Ronning O. Cartilage, function and craniofacial morphogenesis. *Proc Finn Dent Soc* 1991;87:231-7.
  174. Visser NA, Vankampen GPJ, Dekonong MH, Vanderkorst JK. The effects of loading on the synthesis of biglycan and decorin in intact mature articular cartilage in vitro. *Conn Tissue Res* 1994;30:241-50.
  175. Garcia AM, Black AC, Gray ML. Effects of physicochemical factors on the growth of the mandibular condyle in vitro. *Calcif Tissue Int* 1994;54:499-504.
  176. Persson M. The role of movements in the development of sutural and diarthrodial joints tested by long-term paralysis of chick embryos. *J Anat* 1933;137:591-9.
  177. Batmanabane M. Whether mobility influences osteometric features at the articular ends of the metacarpal bones. *Acta Morphol Neerl-Scand* 1982;20:111-5.
  178. Engelsma SO, Janssen HWB, Duterloo HS. An in-vivo transplantation study of the growth of the mandibular condyle in a functional position in the rat. *Arch Oral Biol* 1980;25:305-11.
  179. Price JS, Jackson B, Eastell R, Russell RG, Lanyon LE. The response of the skeleton to physical training: a biochemical study in horses. *Bone* 1995;17:221-7.
  180. Critchlow MA, Bland YS, Ashhurst DE. The effects of age on the response of rabbit periosteal osteoprogenitor cells to exogenous transforming growth factor-(beta)2. *J Cell Sci* 1994;107:499-516.
  181. Voytik SL, Przyborski M, Badlak SF, Konieczny S. Differential expression of muscle regulatory factor genes in normal and denervated adult rat hindlimb muscles. *Dev Dyn* 1993;198:214-24.
  182. Jarvis JC, Sutherland H, Mayne CN, Gilroy SJ, Salmons S. Induction of fast-oxidative phenotype by chronic muscle stimulation: mechanical and biochemical studies. *Am J Physiol* 1996;270:C306-12.
  183. Mayne CN, Sutherland H, Jarvis JC, Gilroy SJ, Craven AJ, Salmons S. Induction of a fast-oxidative phenotype by chronic muscle stimulation: histochemical and metabolic studies. *Am J Physiol* 1996;270:C313-20.
  184. Rouaud T, Fontaine-Perus J, Gardahaut MF. Seasonal variation in the phenotype of the adult ferret cremaster muscle. *Experientia* 1996;52:184-7.

185. Moss ML. An introduction to the neurobiology of orofacial growth. *Acta Biotheor* 1972;22:236-59.
186. Moss ML. Neurotrophic regulation of craniofacial growth. In: McNamara JA, editor. *Control mechanisms of craniofacial growth*. Ann Arbor: University of Michigan Press, 1975: Monogr 3: 25-50.
187. Walker DW, Luff AR. Functional development of fetal limb muscles: a review of the roles of activity, nerves and hormones. *Reprod Fertil Dev* 1995;7:391-8.
188. Grinnell AD. Dynamics of nerve-muscle interaction in developing and mature neuromuscular junctions. *Physiol Rev* 1965;75:789-834.
189. Cooper DM. Evidence for the mechanisms of exercise modulation of growth—an overview. *Med Sci Sports Exerc* 1994;26:733-40.
190. Rafuse VF, Landmesser LT. Contractile activity regulates isoform expression and polysialization of NCAM in cultured myotubes: involvement of  $Ca^{2+}$  and protein kinase C. *J Cell Biol* 1996;132:969-83.
191. Kawakami Y, Abe T, Kuno S-Y, Fukunaga T. Training-induced changes in muscle architecture and specific tension. *Eur J Appl Physiol* 1995;72:37-43.
192. Duchateau J. Bed rest induces neural and contractile adaptations in triceps surae. *Med Sci Sports Exerc* 1995;27:1581-9.
193. Kannus P, Sievanen H, Vuori I. Physical loading, exercise and bone. *Bone* 1996;18(Suppl):1S-3S.
194. Hess B, Mikhailov A. Self-organization in living cells. *Science* 1994;264:223-4.
195. Boyd CAR, Noble D, editors. *The logic of life. The challenge of integrative physiology*. Oxford: Oxford University Press, 1993.
196. Bak P, Paczuski M. Complexity, contingency and criticality. *Proc Natl Acad Sci USA* 1995;92:6689-96.
197. Gunther R, Shapiro B, Wagner P. Complex systems, complexity measures, grammars and model inferring. *Chaos Solitons and Fractals*, 1994;4:635-51.
198. Shea MC. Complexity and evolution: what everybody knows. *Biol Phil* 1991;6: 302-24.
199. Casti JL. *Connectivity, complexity and catastrophe*. New York: John Wiley, 1979.
200. Stein AA. Self-organization in biological systems as a result of interaction between active and passive mechanical stresses: mathematical model. In: Akkas N, editor. *Biomechanics of active movement and division of cells*. Berlin: Springer Verlag, 1994: NATO ASI Series 484:459-64.
201. Mendel JM. Fuzzy logic systems for engineering: a tutorial. *Proc IEEE* 1995;83: 345-77.
202. Corbit JD, Garbarby DJ. Fractal dimension as a quantitative measure of complexity in plant development. *Proc R Soc Lond B Biol Sci* 1995;262:1-6.
203. Yates FE. Order and complexity in dynamical systems—hemodynamics as a generalised mechanics for biology. *Math Comput Model* 1994;19:49-74.
204. Crutchfield JP. Observing complexity and the complexity of observation. In: Atmanspracher H, editor. *Inside versus outside*. Berlin: Springer Verlag, 1993:235-72.
205. Haken H. *Information and self-organization: a macroscopic approach to complex systems*. Berlin: Springer Verlag, 1988.
206. Badii R. Complexity and unpredictable scaling of hierarchical structures. In: Bountis T, editor. *Chaotic dynamics, theory and practice*. New York: Plenum Press, 1994.
207. Grassberger P. Information and complexity measures in dynamical systems. In: Atmanspracher H, Scheingraber H, editors. *Information dynamics*. New York: Plenum Press, 1991.
208. Gell-Mann M. *The Quark and the Jaguar—adventures in the simple and the complex*. London: Little, Brown & Co, 1994.
209. Bonner JT. *The evolution of complexity*. Princeton: Princeton University Press, 1988.
210. Crutchfield JP. Is anything ever new? Considering emergence. In: Cowan G, Pines D, Melzner D, editors. *Complexity: metaphors, models and reality*. Redwood City: Addison-Wesley, 1994:479-97.
211. Mitchell M, Hraber PT, Crutchfield JP. Revisiting the edge of chaos: evolving cellular automata to perform computations. *Complex Systems* 1993;7:89-130.
212. Sipper M. Studying artificial life using a simple, general cellular model. *Artif Life J* 1995;1:1-35.
213. Moss ML. Finite element method comparison of murine mandibular form differences. *J Craniofac Genet Dev Biol* 1988;8:3-20.
214. Moss, ML, Skalak R, Patel H, Sen K, Moss-Salentijn L, Shinozuka M, Vilmann H. Finite element modeling of craniofacial growth. *Am J Orthod* 1985;87:453-72.
215. Cheverud JM, Hartman SE, Richtsmeier JT, Atchley WR. A quantitative genetic analysis of localized morphology in mandibles of inbred mice using finite element scaling analysis. *J Craniofac Genet Dev Biol* 1991;11:122-37.
216. McGuinness N, Wilson AN, Jones M, Middleton J, Robertson NR. Stresses induced by edgewise appliances in the periodontal ligament—a finite element study. *Angle Orthod* 1992;62:15-22.
217. Hart RT, Hennebel VV, Thongpreda N, VanBuskirk WC, Anderson RC. Modeling the biomechanics of the mandible: a three-dimensional finite element study. *J Biomech* 1992;25:261-86.
218. Lele S, Richtsmeier JT. On comparing biological shapes: detection of influential landmarks. *Am J Phys Anthropol* 1992;87:49-65.
219. Fine MB, Lavelle CLB. Diagnosis of skeletal form on the lateral cephalogram with a finite element-based expert system. *Am J Orthod Dentofac Orthop* 1992;101:318-29.
220. Richtsmeier JT. Comparative study of normal, Cruzon and Apert craniofacial morphology using finite element scaling analysis. *Am J Phys Anthropol* 1987;74: 473-93.
221. Koriath TWP, Romilly DP, Hannam AG. Three-dimensional finite element stress analysis of the dentate human mandible. *Am J Phys Anthropol* 1992;88:69-96.
222. Molenaar PCM, Boomsma DI, Dolan CV. A third source of developmental differences. *Behav Genet* 1993;23:519-24.